

# Advances in Uveal Melanoma: From Molecular Pathogenesis to Precision Diagnostics and Personalized Therapies: Narrative Overview

Raed Shatnawi<sup>1</sup>, Ahmad Al-Hyari<sup>2</sup>, Motasem Al-Latayfeh<sup>1</sup>, Mohammad Abu Ain<sup>1</sup>, Husam Shatnawi<sup>3</sup>, Yazan Shatnawi<sup>3</sup>, Yacoub A. Yousef<sup>4</sup>

<sup>1</sup>Department of Special Surgery, Faculty of Medicine, The Hashemite University, P.O. Box 330127, Zarqa 13133, Jordan.

<sup>2</sup>Department of Ophthalmology, Prince Hamzah Hospital, Ministry of Health, Amman, Jordan. <sup>3</sup>Medical Student, Faculty of Medicine, The Hashemite University, P.O. Box 330127, Zarqa 13133, Jordan. <sup>4</sup>Department of Surgery, King Hussein Cancer Center, Amman, Jordan.

## Abstract

**Objective:** This review aims to summarize recent progress in the molecular understanding, diagnostic strategies, and treatment innovations in uveal melanoma (UM), the most common primary intraocular malignancy in adults. Emphasis is placed on the integration of precision diagnostics and emerging therapies that may improve clinical outcomes in high-risk cases. **Materials and Methods:** A narrative literature review was conducted using databases including PubMed, Scopus, Web of Science, and Google Scholar, covering the years 2020 to 2024. Keywords used included “uveal melanoma,” “liquid biopsy,” “circulating tumor cells,” “gene mutations,” “immunotherapy,” and “precision oncology.” Relevant peer-reviewed articles, clinical trials, and reviews were selected based on methodological quality and relevance to the scope of the review. **Results:** Uveal melanoma most frequently arises in the choroid and is genetically distinct from cutaneous melanoma. It is primarily driven by mutations in guanine nucleotide-binding protein G (q) subunit alpha (GNAQ), guanine nucleotide-binding protein G(q) subunit alpha-11 (GNA11), BRCA1 associated protein-1 (BAP1), eukaryotic translation initiation factor 1A X-linked (EIF1AX), and splicing factor 3B subunit 1 (SF3B1). These mutations activate key signaling pathways such as mitogen-activated protein kinase (MAPK) and phosphoinositide 3-kinase/protein kinase B (PI3K/AKT), influencing prognosis and therapeutic response. Diagnostic advancements include high-resolution imaging and liquid biopsy techniques, which enable detection of circulating tumor cells, circulating tumor DNA, and microRNAs. Standard treatments include radiation therapy (plaque brachytherapy) and surgical interventions. Novel therapeutic approaches such as tebentafusp (a T-cell receptor therapy), oncolytic viruses, chimeric antigen receptor (CAR) T-cell therapy, suicide gene constructs, and RNA interference show promise in clinical and preclinical settings. **Conclusion:** A precision medicine approach that integrates molecular diagnostics, artificial intelligence-enhanced liquid biopsy, and novel systemic therapies is transforming the management of uveal melanoma. These innovations may enable earlier detection, more accurate risk stratification, and targeted treatment, potentially improving survival and preserving vision in affected patients.

**Keywords:** Uveal melanoma- liquid biopsy- circulating tumor cell- GNAQ mutation- tebentafusp- RNA interference

*Asian Pac J Cancer Biol*, **10 (3)**, 759-768

Submission Date: 03/23/2025      Acceptance Date: 05/27/2025

## Introduction

Uveal melanoma (UM) is the most prevalent form of primary intraocular malignancy in adults, although it remains a rare disease with an incidence of approximately 5 cases per million annually in the United States [1, 2]. Despite its rarity, UM carries a significant

clinical burden due to its aggressive metastatic potential. In a population-based study from Sweden, about 30% of patients died from metastatic UM within five years of diagnosis, with mortality rising to nearly 40% over a span of 10 to 15 years. The liver and gastrointestinal

## Corresponding Author:

Dr. Raed Shatnawi

Associate Professor of Ophthalmology, Department of Special Surgery, Faculty of Medicine, The Hashemite University, Zarqa 13133, Jordan.

Email: raed972@yahoo.com

tract, particularly the colon, are the most common sites of metastasis and primary contributors to mortality [1].

Beyond its biological aggressiveness, UM severely impacts patients' emotional well-being and mental health. The diagnosis often leads to significant psychological distress, including anxiety, depression, and a diminished quality of life, especially given the associated risks of visual impairment and systemic spread [3]. Although therapeutic innovations such as tebentafusp and immune-modulating T-cell receptor (TCR) therapy have been introduced, they have not markedly improved overall survival outcomes in patients with metastatic disease [4]. One reason for this is the likely presence of micrometastases at the time of diagnosis, suggesting early hematogenous dissemination. Additionally, diagnostic and treatment delays are thought to exacerbate disease progression and worsen prognoses [5].

UM originates from melanocytes in the uveal tract of the eye, including the iris, ciliary body, and choroid [6]. It is genetically distinct from cutaneous melanoma, being driven by mutations in specific genes such as BRCA1-associated protein 1 (BAP1), eukaryotic translation initiation factor 1A X-linked (EIF1AX), guanine nucleotide-binding protein G (q) subunit alpha (GNAQ), guanine nucleotide-binding protein G (q) subunit alpha-11 (GNA11), and splicing factor 3B subunit 1 (SF3B1) [7]. These genetic alterations influence tumor development and metastatic behavior. For example, BAP1 loss is linked with poor prognosis and high metastatic potential, whereas EIF1AX mutations tend to indicate a more favorable outcome. Mutations in GNAQ and GNA11 activate signaling cascades such as RAS and phosphoinositide 3-kinase (PI3K), promoting tumor proliferation, survival, and migration [8].

Recent discoveries have identified circulating hybrid cells (CHCs), which form via fusion of tumor and immune cells and carry markers from both, making them promising indicators of metastatic risk [9]. Likewise, tumor-derived extracellular vesicles (TEVs) are believed to contribute to metastasis by altering distant tissue microenvironments [10].

The discovery of CHCs and TEVs in blood has sparked interest in liquid biopsy technologies. These non-invasive tests detect circulating tumor cells (CTCs), cell-free DNA (cfDNA), and exosomes, providing real-time insights into tumor dynamics. Liquid biopsies allow for early detection of metastasis, continuous monitoring of disease progression, and evaluation of therapeutic response all without the need for tissue samples [1]. When paired with artificial intelligence (AI) systems, data from liquid biopsies can improve diagnostic precision, guide treatment decisions, and support personalized patient care.

This review will comprehensively explore UM's molecular basis, key genetic mutations, metastatic mechanisms, diagnostic innovations, and therapeutic challenges, with emphasis on integrating liquid biopsy and artificial intelligence to enhance clinical outcomes. This review advances precision medicine by summarizing molecular insights, non-invasive diagnostics, and emerging therapies that support early detection, personalized

risk assessment, and targeted treatment. It also aligns with the efforts to improve clinical outcomes through cost-effective technologies, innovative procedures, and culturally adapted care strategies, offering a foundation for future research and policy development as other studies confirmed [11-17].

## Methods

This narrative literature review was conducted to synthesize current knowledge on the pathogenesis, diagnosis, and management of uveal melanoma (UM), with a particular focus on emerging diagnostic technologies such as liquid biopsy and artificial intelligence (AI)-assisted tools. A systematic literature search was performed using multiple academic databases, including PubMed, Scopus, Web of Science, and Google Scholar, covering publications from 2020 to 2024. Search terms included combinations of keywords such as "uveal melanoma," "choroidal melanoma," "intraocular tumor," "liquid biopsy," "circulating tumor cells," "circulating tumor DNA," "microRNA," "prognostic biomarkers," "ocular oncology," "plaque brachytherapy," and "artificial intelligence in ocular diagnosis."

Relevant peer-reviewed articles, clinical trials, meta-analyses, and systematic reviews were included based on their scientific quality, clinical relevance, and publication recency. Preference was given to studies providing molecular insights, epidemiological trends, diagnostic innovations, and therapeutic efficacy in the context of UM. Reference lists of the most impactful articles were manually screened to identify additional studies that were not retrieved through initial database searches.

Only publications in the English language were considered for inclusion. Extracted data were organized thematically into five major domains: (1) molecular pathophysiology and genetic underpinnings; (2) epidemiology and clinical risk stratification; (3) diagnostic modalities, including advanced imaging and tissue-based assessment; (4) novel diagnostic innovations such as liquid biopsy and circulating biomarkers; and (5) contemporary and experimental therapeutic interventions, including surgery, radiation, immunotherapy, gene therapy, and AI-assisted tools.

Each identified article was assessed for its contribution to understanding the pathophysiology, diagnostic advancements, and therapeutic landscape of UM, particularly in relation to circulating biomarkers and digital diagnostic tools. Studies specifically addressing liquid biopsy techniques such as detection of circulating tumor cells (CTCs), circulating tumor DNA (ctDNA), microRNAs (miRNAs), and tumor-derived extracellular vesicles (TEVs) were prioritized to explore their utility in early detection, risk stratification, and treatment monitoring. Articles detailing the development and validation of AI-enabled algorithms for image analysis or molecular data interpretation in ocular oncology were also included.

Emphasis was placed on integrating findings across multiple disciplines, including molecular oncology,

ophthalmology, bioinformatics, and biomedical engineering. The collected data were synthesized qualitatively, focusing on the convergence of histopathological findings, genetic profiling, and non-invasive diagnostic technologies to form a comprehensive understanding of UM's clinical and biological behavior.

## Results

### *Pathophysiology*

Uveal melanoma (UM) originates from melanocytes residing in the uveal tract, which includes the choroid, ciliary body, and iris. Approximately 90% of cases arise in the choroid [1, 18]. This malignancy is characterized by marked genetic instability and a high tendency for hematogenous metastasis, particularly to the liver, which contributes significantly to its poor long-term prognosis even when the primary tumor is controlled. Unlike cutaneous melanoma, UM is not strongly associated with ultraviolet (UV) radiation exposure. Instead, established risk factors include fair skin, light-colored eyes, atypical nevi, ocular melanocytosis (also known as nevus of Ota), and periorbital freckling [2, 19]. Certain occupational exposures such as welding or working in high-heat environments have also been implicated in increasing the risk of UM.

On a molecular level, UM is driven by mutations that differ from those found in cutaneous melanoma. Activating mutations in guanine nucleotide-binding protein G (q) subunit alpha (GNAQ) and guanine nucleotide-binding protein G (q) subunit alpha-11 (GNA11) initiate tumorigenic signaling cascades including the mitogen-activated protein kinase (MAPK), phosphoinositide 3-kinase/protein kinase B (PI3K/AKT), and mammalian target of rapamycin (mTOR) pathways, all of which promote unchecked cellular proliferation and survival. Loss of function in BRCA1-associated protein 1 (BAP1) is a particularly aggressive molecular alteration, linked to early metastasis and poor survival outcomes. In contrast, mutations in eukaryotic translation initiation factor 1A X-linked (EIF1AX) and splicing factor 3B subunit 1 (SF3B1) are generally associated with a more indolent disease course and improved prognosis [5, 20].

UM arises in the immune-privileged environment of the eye a setting evolved to minimize inflammation and preserve vision. However, malignancy disrupts this immune balance. The tumor microenvironment in UM is often marked by an influx of macrophages and lymphocytes and an upregulation of human leukocyte antigen (HLA) class I and II molecules. This immune response is further amplified by activation of the nuclear factor kappa-light-chain-enhancer of activated B cells (NFκB) pathway, which supports tumor survival and progression [1, 3].

Recent findings have expanded the understanding of UM progression through mechanisms involving tumor-derived extracellular vesicles (TEVs) and circulating hybrid cells (CHCs). TEVs are membrane-bound vesicles secreted by tumor cells that carry oncogenic

proteins including MAPK pathway components, vascular endothelial growth factor (VEGF), and Wnt modulators facilitating the creation of pre-metastatic niches, especially in the liver. CHCs, formed through the fusion of tumor cells with immune cells, express both immune and tumor markers and possess high tumorigenic potential. The detection of TEVs and CHCs in bodily fluids suggests their potential role as non-invasive biomarkers in early diagnosis and monitoring of UM via liquid biopsy [18-27].

### *Epidemiology*

Uveal melanoma (UM) is the most common primary intraocular malignancy in adults, though it remains rare, with an incidence of approximately 5 cases per million annually in the United States [1,2]. Several classification systems have been developed to aid in clinical staging, prognostication, and treatment planning. The American Joint Committee on Cancer Tumor-Node-Metastasis (AJCC TNM) system is widely used, assessing tumor size (T), invasion into ocular structures, lymph node involvement (N), and the presence of distant metastasis (M) [1, 23]. Another key framework is the Collaborative Ocular Melanoma Study (COMS) classification, which stratifies tumors into small, medium, or large categories based on diameter and thickness critical for both clinical trials and therapeutic decisions. Histologically, the modified Callender system categorizes tumors as spindle-cell, epithelioid-cell, or mixed-cell types, each bearing distinct prognostic implications.

Recent advances in molecular diagnostics have enhanced risk stratification through tools such as gene expression profiling (GEP) and The Cancer Genome Atlas (TCGA)-based classification. These techniques help identify genetic signatures associated with high metastatic risk, enabling more personalized follow-up and therapeutic approaches [1, 24].

Clinically, UM is often asymptomatic in early stages and frequently discovered incidentally during routine ophthalmologic evaluations. When symptoms do occur, they are usually dictated by tumor location and may include reduced visual acuity, photopsia (flashes of light), floaters, metamorphopsia (distorted vision), or localized field defects [1, 3, 24, 25]. Iris tumors may present with visible pigment changes or mass lesions and are detected earlier due to their anterior location. In contrast, melanomas of the ciliary body often remain undiagnosed until they are large, due to their posterior and hidden position within the eye. Diagnostic delays are common, with approximately one-quarter of symptomatic cases initially misidentified or overlooked [1, 3, 24].

Prognosis in UM is influenced by anatomical, histopathological, and genetic factors. Tumors located in the ciliary body or near the optic nerve tend to be more aggressive and harder to detect early, leading to poorer outcomes. Histopathological markers of poor prognosis include epithelioid cell type, high mitotic rate, and extrascleral extension. Cytogenetically, monosomy 3, gain of chromosome 8q, and loss of chromosome 6q are associated with a high risk of metastasis, whereas gain of chromosome 6p is considered a favorable prognostic

indicator [1, 5, 24–30].

To differentiate early melanomas from benign nevi, clinicians often use the mnemonic TFSOM-UHHD, which stands for: Thickness >2 mm, subretinal Fluid, Symptoms, Orange pigment, Margin near optic disc, Ultrasonographic Hollowness, Halo absence, and Drusen absence. This tool assists in early detection and timely intervention of malignant lesions [1, 5, 30].

### Diagnosis

The diagnosis of uveal melanoma (UM) requires a multimodal approach that integrates clinical evaluation, advanced imaging techniques, and, when necessary, histopathological confirmation. The process often begins with a detailed dilated fundus examination, which may reveal a characteristic pigmented or dome-shaped lesion. In advanced cases, the lesion may adopt a mushroom or collar-button configuration due to rupture of Bruch's membrane a hallmark feature distinguishing UM from benign intraocular lesions [1, 3-5, 31].

Optical coherence tomography (OCT) is a critical non-invasive imaging modality that provides high-resolution cross-sectional images of the retina and choroid. OCT is particularly useful in detecting subretinal fluid, disruption of the retinal pigment epithelium (RPE), photoreceptor loss, and retinoschisis. These features aid in differentiating UM from benign entities such as choroidal nevi or vascular tumors like hemangiomas [25-32].

Ocular ultrasonography remains an essential diagnostic tool, especially when direct fundus visualization is obstructed by media opacities. A-scan and B-scan ultrasonography allow accurate measurement of tumor thickness and internal reflectivity. UM typically exhibits low to medium reflectivity, dome or mushroom-shaped morphology, and may be associated with choroidal excavation and exudative retinal detachment [1].

Magnetic resonance imaging (MRI) is particularly valuable for assessing posteriorly located or large tumors and for evaluating potential extrascleral extension. UM lesions usually appear hyperintense on T1-weighted and hypointense on T2-weighted MRI sequences. Restricted diffusion may also be seen and is indicative of high cellularity, supporting the diagnosis of malignancy [1, 30-35].

Definitive diagnosis is established through histopathological analysis, which may follow fine needle aspiration biopsy (FNAB) or enucleation. Microscopic examination identifies tumor cell type spindle, epithelioid, or mixed with significant prognostic value. Spindle-cell tumors are generally associated with favorable outcomes, while epithelioid-cell tumors suggest a poorer prognosis. Additional histologic markers, such as mitotic index, vascular loop presence, and lymphocytic infiltration, provide further insight into tumor aggressiveness and metastatic potential [1, 25-36].

Ultimately, diagnosis hinges on the integration of clinical, radiologic, and histologic data. Advances in imaging and molecular diagnostics have enhanced the accuracy of UM identification and staging, facilitating earlier and more tailored treatment planning.

### Key Diagnostic Themes and Molecular Targets in Liquid Biopsy

Liquid biopsy has emerged as a transformative, non-invasive tool for the diagnosis and monitoring of uveal melanoma (UM), particularly in settings where traditional tissue sampling is technically challenging or carries procedural risk. While fine needle aspiration biopsy (FNAB) remains useful in select cases, its limitations including potential complications such as retinal detachment, vitreous hemorrhage, and tumor seeding have prompted the exploration of safer alternatives [33-36].

Liquid biopsy involves the analysis of tumor-derived components circulating in bodily fluids such as blood, aqueous humor, and vitreous fluid. This technique enables dynamic monitoring of disease progression and therapeutic response without direct tumor manipulation, supporting its integration into precision oncology workflows [1, 35-38].

Key analytes in liquid biopsy include circulating tumor cells (CTCs), circulating tumor DNA (ctDNA), microRNAs (miRNAs), and extracellular vesicles. CTCs reflect active tumor dissemination and carry genetic mutations consistent with the primary lesion. Their detection often via immunomagnetic separation or filtration correlates with metastatic potential and poorer prognosis [1, 38-40].

ctDNA, shed into the bloodstream by apoptotic or necrotic tumor cells, harbors actionable mutations such as those in guanine nucleotide-binding protein G(q) subunit alpha (GNAQ) and guanine nucleotide-binding protein G (q) subunit alpha-11 (GNA11). Although ctDNA concentrations are typically low in early-stage UM, their presence can guide molecular profiling and longitudinal surveillance [1, 39].

Circulating miRNAs, including miRNA-618, serve as epigenetic biomarkers with differential expression patterns in metastatic versus non-metastatic disease. These small, stable RNA fragments can help stratify patient risk and potentially indicate early progression. Protein biomarkers such as glycoprotein 100 (gp100), osteopontin, cathepsin, and heat shock protein 27 are also under investigation, particularly in multiplex panels to enhance diagnostic sensitivity [35-40].

Extracellular vesicles, especially tumor-derived subtypes, represent another promising class of biomarkers. These vesicles carry oncogenic cargo including MAPK pathway proteins, vascular endothelial growth factor (VEGF), and Wnt modulators and are instrumental in creating pre-metastatic niches, particularly in the liver. Their detection, however, remains technically challenging due to isolation and characterization barriers [34-41].

Advanced platforms such as next-generation sequencing (NGS), enzyme-linked immunosorbent assay (ELISA), fluorescence-activated cell sorting (FACS), and nanoparticle tracking analysis (NTA) are being developed to improve detection accuracy. When combined with artificial intelligence (AI) and integrative molecular analysis, liquid biopsy holds promise for revolutionizing early detection, risk stratification, and personalized treatment strategies in UM.



### *Current Treatment Methods*

Management of uveal melanoma (UM) is largely influenced by tumor size, location, and proximity to critical ocular structures, with the primary goal of achieving local control while preserving as much vision as possible. For anteriorly situated tumors especially those confined to the iris or ciliary body transscleral resection, also known as exo-resection, may be performed. This surgical technique involves the partial or total removal of the tumor through a direct external approach but is generally avoided for posterior tumors due to increased procedural complexity and risk [1, 34, 37-43].

When tumors are large or have caused significant ocular damage, enucleation, the complete surgical removal of the eye, may be indicated. This option is typically reserved for lesions exceeding 12 mm in thickness or 18 mm in basal diameter, or in cases complicated by secondary glaucoma or optic nerve involvement. Although enucleation can provide definitive local control, it does not prevent systemic spread, and recurrence portends a poor prognosis [1, 40-43].

In rare, advanced cases involving extensive orbital invasion or a blind and painful eye, orbital exenteration may be required. This disfiguring procedure entails the removal of the eye along with adjacent orbital contents, and while potentially curative in select cases, it is associated with significant morbidity and limited overall benefit when recurrence occurs postoperatively [1, 40-43].

Radiation therapy, particularly plaque brachytherapy, is a preferred eye-conserving modality for small to medium-sized tumors. This technique involves temporarily attaching a radioactive plaque (commonly iodine-125 or ruthenium-106) to the sclera overlying the tumor, delivering localized radiation. It is most effective for tumors not encroaching upon the optic nerve. Complications may include radiation-induced cataracts, retinopathy, and optic neuropathy, depending on tumor proximity and radiation dose [39-52].

Given the complexity and variability of UM presentations, treatment must be individualized. Decisions are best made within a multidisciplinary framework, considering the patient's overall health, tumor characteristics, visual function, and personal preferences. The aim is to optimize tumor control while minimizing long-term functional impairment and enhancing quality of life [1].

### *Contemporary Therapeutic Modalities*

Contemporary approaches to treating uveal melanoma (UM) are increasingly focused on addressing the limitations of conventional therapies, particularly in the context of metastatic disease, where long-term survival remains poor. One major area of innovation involves immunotherapy, specifically immune checkpoint inhibitors that block cytotoxic T-lymphocyte-associated protein 4 (CTLA-4) and programmed cell death protein 1/programmed death-ligand 1 (PD-1/PD-L1) interactions. Despite their success in other malignancies, agents like ipilimumab and nivolumab have demonstrated limited efficacy in UM due to the tumor's low mutational burden

and an immunosuppressive microenvironment. In contrast, tebentafusp, a T cell receptor (TCR)-based therapy targeting the human leukocyte antigen HLA-A\*0201, has shown improved overall survival in eligible patients, marking a significant step forward in precision immunotherapy [1, 43-53].

Oncolytic virotherapy is another promising avenue. Engineered viruses such as talimogene laherparepvec (T-VEC), herpes simplex virus-1 (HSV-1), and echovirus-7 (ECHO-7) are designed to selectively infect and lyse malignant cells while stimulating immune responses. Notably, AU-011 (belzupacap sarotalocan), a virus-like drug conjugate, has demonstrated encouraging outcomes in clinical trials for local tumor control and vision preservation and is progressing into advanced trial phases [47-53].

Adoptive T cell therapy, including tumor-infiltrating lymphocytes (TILs) and chimeric antigen receptor (CAR) T cells, offers an individualized immunologic strategy for patients with metastatic UM unresponsive to other treatments. Preclinical and early-phase clinical data suggest that CAR T cells can mediate tumor regression by targeting specific antigens, potentially overcoming resistance mechanisms inherent to UM [44-49].

Gene therapy strategies are also under development. Suicide gene therapy, for example, introduces enzymes such as cytosine deaminase into tumor cells to convert prodrugs into cytotoxic agents directly at the tumor site. Additionally, constructs targeting B7-H3 a surface antigen overexpressed in UM using inducible CRISPR-associated protein 9 (iCas9) systems have shown efficacy in reducing metastasis in animal models, offering a precision-guided treatment platform [1, 48-53].

RNA-based therapies are advancing rapidly, particularly RNA interference (RNAi) strategies using small interfering RNAs (siRNAs) and microRNAs (miRNAs) to silence key oncogenic drivers like vascular endothelial growth factor (VEGF), B-cell lymphoma 2 (Bcl-2), and hypoxia-inducible factor 1-alpha (HIF-1α). The use of innovative delivery systems, such as hyaluronic acid (HA)-coated chitosan nanoparticles, is being explored to enhance therapeutic stability and cellular uptake. Long non-coding RNAs (lncRNAs), including miR-181a, have also been implicated as potential therapeutic targets, although clinical translation is challenged by issues of delivery and degradation [1, 49-53].

Collectively, these evolving therapeutic strategies reflect a shift toward highly tailored treatment paradigms in UM. By combining immunological, genetic, and molecular techniques, ongoing research aims to address the unmet need for effective systemic treatments in a cancer historically resistant to conventional approaches.

## **Discussion**

Recent advances in the understanding of uveal melanoma (UM) have significantly enriched our ability to diagnose, stratify, and potentially treat this aggressive intraocular malignancy. Although UM remains a rare entity with a relatively low incidence, its high metastatic

potential and liver tropism necessitate early detection and effective management strategies [1-3]. The observed five- to ten-year mortality rates reaching 40% underscore the urgency of refining both systemic surveillance and therapeutic options [4-6]. These epidemiologic realities highlight the importance of molecular profiling and personalized interventions aimed at intercepting disease progression at earlier stages.

At the molecular level, UM is now recognized as genetically distinct from cutaneous melanoma, largely due to its unique set of driver mutations. Mutations in *GNAQ* and *GNA11* genes initiate aberrant signaling through the MAPK and PI3K/AKT pathways, contributing to cell proliferation and survival [22–27,29–31]. Importantly, the loss of *BAP1* function serves as a hallmark of poor prognosis and increased metastatic risk, while mutations in *EIF1AX* and *SF3B1* appear to signal less aggressive tumor behavior [33-38]. These genetic markers not only assist in prognostication but also serve as potential therapeutic targets, guiding risk-adapted surveillance and clinical decision-making.

The tumor microenvironment in UM is shaped by immune privilege, a physiological state intended to preserve ocular function by limiting inflammation. However, this privilege may inadvertently favor tumor immune evasion. Studies have shown that inflammatory UM phenotypes characterized by lymphocytic infiltration and elevated HLA class I/II expression are paradoxically associated with worse outcomes [28-37]. The activation of NF $\kappa$ B signaling in both primary tumors and metastases supports a pro-survival and pro-metastatic state, reinforcing the idea that immunomodulatory strategies need to be tailored to this unique immune context [52-57].

Liquid biopsy technologies offer a promising path forward in early UM detection and longitudinal disease monitoring. Detection of circulating tumor cells (CTCs), circulating tumor DNA (ctDNA), and microRNAs (miRNAs) in blood and ocular fluids circumvents the limitations of fine needle aspiration biopsy, which carries risks of retinal detachment and tumor seeding [39-44]. Importantly, CTCs have demonstrated prognostic value, with high counts correlating with reduced progression-free and overall survival [22-25]. Moreover, studies confirm the presence of shared copy number alterations between CTCs and primary tumors, suggesting their utility for non-invasive genotyping and molecular tracking [1].

Circulating tumor DNA further refines molecular profiling, with next-generation sequencing of ctDNA enabling the identification of *GNAQ* and *GNA11* mutations, as well as the monitoring of treatment response [30-35]. Despite challenges in detecting ctDNA in early-stage disease due to low abundance, advancements in isolation techniques continue to improve analytical sensitivity. Meanwhile, miRNAs such as miRNA-618 and other stable blood-based biomarkers offer additional layers of specificity for identifying metastatic risk [19, 28-35]. Together, these analytes strengthen the case for routine liquid biopsy integration into UM clinical protocols.

Current standard-of-care treatments, including transscleral resection, enucleation, exenteration, and

plaque brachytherapy, offer varying degrees of disease control and ocular preservation depending on tumor size and location [24-32]. While effective for localized disease, these approaches fail to address systemic metastasis, particularly in the liver. This limitation reinforces the need for systemic therapies capable of eradicating micrometastatic disease or modulating immune response to prevent metastatic outgrowth.

Immunotherapy has emerged as a key investigational avenue but has shown modest efficacy in UM compared to cutaneous melanoma. Checkpoint inhibitors such as nivolumab and ipilimumab have yielded limited objective response rates, likely due to the low mutational burden of UM and its immunosuppressive microenvironment [34-46]. However, tebentafusp has demonstrated survival benefits in HLA-A\*0201-positive patients, suggesting the need for genotype-based patient selection and expansion of HLA-compatible immunotherapies [38-52].

Gene-directed therapies, including suicide gene therapy and RNA interference strategies, represent additional innovative modalities under preclinical and clinical evaluation. Studies involving cytosine deaminase constructs and targeted siRNA delivery have shown success in suppressing tumor proliferation in vitro and in animal models [39-47]. However, clinical translation remains limited by challenges in vector design, delivery efficiency, and off-target effects. The potential for B7-H3-targeted CAR-T constructs and lncRNA-based therapies also holds promise, though these require further validation in larger, controlled trials [37-43].

Adoptive cell therapy (ACT), including tumor-infiltrating lymphocytes (TILs) and CAR-T cells, has demonstrated capacity to induce tumor regression in select metastatic UM cases [49-53]. Notably, engineered CAR-T cells have exhibited in vivo efficacy in murine models, including resistance to conventional TIL-based therapies. These findings support the rationale for integrating ACT into treatment regimens, particularly for patients refractory to immune checkpoint inhibitors or those with high-risk molecular profiles.

These scientific developments in the field of uveal melanoma (UM), particularly those related to molecular diagnostics, liquid biopsy, and novel immunotherapeutic strategies, are emblematic of a broader shift toward precision and patient-centered care in oncology. However, technological advancements alone are insufficient to optimize patient outcomes unless they are supported by comprehensive institutional frameworks that prioritize safety, quality, and individualized care. For instance, the integration of AI-assisted diagnostic tools and real-time monitoring systems must be matched with enhanced protocols for fall risk screening and management especially in patients with visual impairments or those undergoing aggressive treatments like enucleation or radiation therapy [1, 51]. Vision loss and balance issues place UM patients at higher risk of inpatient falls, necessitating proactive screening, tailored safety plans, and environmental modifications to prevent avoidable harm. Similarly, ensuring structured continuity of care after discharge through coordinated follow-up

appointments, medication reconciliation, and symptom surveillance helps detect recurrence or metastasis early, mitigates treatment-related complications, and reinforces patient adherence to long-term care plans [52, 53].

Moreover, to truly elevate the quality of care, institutions must prioritize patient-centered models that emphasize education, psychosocial support, and shared decision-making. UM patients, who often confront irreversible vision loss and the psychological toll of cancer diagnosis, benefit substantially from structured educational interventions that clarify treatment options, potential side effects, and self-monitoring strategies [54-58]. Incorporating mental health services, peer support groups, and low-vision rehabilitation into the treatment ecosystem addresses both emotional resilience and functional recovery. Finally, systemic quality improvement strategies aimed at reducing diagnostic and procedural errors such as the use of checklists, standardized documentation, and continuous process audits are essential for sustaining institutional accountability and patient safety [58-68]. These reforms underscore the need for advance multidisciplinary integration and management practice involving oncologists, ophthalmologists, radiologists, pathologists, nurses, and IT professionals, all working in concert to ensure that scientific innovation translates into meaningful clinical impact [68-78].

In conclusion, the landscape of UM management is rapidly transitioning from static, anatomy-based approaches to dynamic, personalized strategies driven by molecular diagnostics and targeted therapeutics. Liquid biopsy, AI-assisted diagnostics, immune checkpoint modulation, and gene therapies are reshaping how clinicians understand, monitor, and treat UM. Continued multidisciplinary collaboration, coupled with access to multicenter clinical trials and real-world data, will be essential in transforming these promising innovations into clinical standards. Future directions should emphasize individualized treatment based on genetic and biomarker profiling to improve both survival and quality of life for patients affected by this rare but deadly malignancy.

## Acknowledgments

### *Statement of Transparency and Principals*

- Author declares no conflict of interest
- Study was approved by Research Ethic Committee of author affiliated Institute.
- Study's data is available upon a reasonable request.
- All authors have contributed to implementation of this research.

## References

- Kulbay M, Marcotte E, Remtulla R, Lau THA, Paez-Escamilla M, Wu KY, Burnier MN. Uveal Melanoma: Comprehensive Review of Its Pathophysiology, Diagnosis, Treatment, and Future Perspectives. *Biomedicine*. 2024 08 05;12(8):1758. <https://doi.org/10.3390/biomedicine12081758>
- Huang Y, Guo Y. Quality of life among people with eye cancer: a systematic review from 2012 to 2022. *Health and Quality of Life Outcomes*. 2024 01 07;22(1):3. <https://doi.org/10.1186/s12955-023-02219-6>
- Stålhammar G. Comprehensive causes of death in uveal melanoma: mortality in 1530 consecutively diagnosed patients followed until death. *JNCI cancer spectrum*. 2023 Oct 31;7(6):pkad097. <https://doi.org/10.1093/jncics/pkad097>
- Barker CA, Kozlova A, Shoushtari AN, Hay JL, Francis JH, Abramson DH. Quality of Life Concerns in Patients with Uveal Melanoma after Initial Diagnosis. *Ocular Oncology and Pathology*. 2020 05;6(3):184-195. <https://doi.org/10.1159/000502549>
- Stålhammar G. Delays between uveal melanoma diagnosis and treatment increase the risk of metastatic death. *Ophthalmology*. 2023;13(9):1094-104. <https://doi.org/10.1016/j.ophtha.2023.11.021>
- Barbagallo C, Stella M, Broggi G, Russo A, Caltabiano R, Ragusa M. Genetics and RNA Regulation of Uveal Melanoma. *Cancers*. 2023 01 26;15(3):775. <https://doi.org/10.3390/cancers15030775>
- Silva-Rodríguez P, Fernández-Díaz D, Bande M, Pardo M, Loidi L, Blanco-Teijeiro MJ. GNAQ and GNA11 Genes: A Comprehensive Review on Oncogenesis, Prognosis and Therapeutic Opportunities in Uveal Melanoma. *Cancers*. 2022 06 22;14(13):3066. <https://doi.org/10.3390/cancers14133066>
- Dietz MS, Sutton TL, Walker BS, Gast CE, Zarour L, Sengupta SK, Swain JR, et al. Relevance of circulating hybrid cells as a non-invasive biomarker for myriad solid tumors. *Scientific Reports*. 2021 07 01;11(1):13630. <https://doi.org/10.1038/s41598-021-93053-7>
- Tsering T, Laskaris A, Abdouh M, Bustamante P, Parent S, Jin E, Ferrier ST, et al. Uveal Melanoma-Derived Extracellular Vesicles Display Transforming Potential and Carry Protein Cargo Involved in Metastatic Niche Preparation. *Cancers*. 2020 Oct 11;12(10):2923. <https://doi.org/10.3390/cancers12102923>
- Jin E, Burnier JV. Liquid Biopsy in Uveal Melanoma: Are We There Yet?. *Ocular Oncology and Pathology*. 2021 03;7(1):1-16. <https://doi.org/10.1159/000508613>
- Ayaad O, Al Ajmi AA, Al Baimani K, Alhajjiaa EA, Ibrahim R, AlDhahli SN, et al. Breast cancer awareness, screening practices, barriers, and educational interventions in Middle Eastern countries: Challenges and successes. *Asian Pacific Journal of Cancer Biology*. 2025;10(2):251-60. <https://doi.org/10.31557/apjcb.2025.10.2.251-260>
- Ayyad M, Ayaad O, Alkhatatbeh H, Qaddumi B, Sawaqed F, Al-Rawashdeh S. Flexible cystodiathermy for the treatment of recurrent superficial bladder transitional cell carcinoma; efficacy, safety, and cost-effectiveness. *Journal of Renal Injury Prevention*. 2025 02 25;14(4):e38386-e38386. <https://doi.org/10.34172/jrip.2025.38386>
- Ayyad M, Ayaad O, Qaddumi B, Al-Rawashdeh S, Alkhatatbeh H, Al-Baimani K, Ibrahim R, et al. Epidemiological Analysis of Prostatic Cancer: Incidence, Prevalence, Mortality, and Disability Burden in Middle Eastern Countries. *Asian Pacific Journal of Cancer Biology*. 2025 05 04;10(2):393-400. <https://doi.org/10.31557/apjcb.2025.10.2.393-400>
- Ayyad M, Zahra KA, Al Demour S, Al-Zubi MT, Ayaad O. Longitudinal assessment of clinical and functional outcomes following thulium laser enucleation of the prostate in Jordan. *Journal of Lasers in Medical Sciences*. 2025;16: e11.
- Al Ajmi A, Ayaad O, Al Aghbari S, Al Balushi MN, Koziha EK, Al-Ishaq Z, et al. Young women's breast cancer in Gulf countries. *Journal of Young Women's Breast Cancer Health*. 2024;1(1&2):36-41. [https://doi.org/10.4103/YWBC.YWBC\\_14\\_24](https://doi.org/10.4103/YWBC.YWBC_14_24)
- Ayyad M, Ayaad O, Alkhatatbeh H, Sawaqed F, Al-



- Rawashdeh S, Qaddumi B. Gabapentin for overactive bladder: A quasi-experimental study. *Immunopathologia Persa*. 2023;10(1):e40574. <https://doi.org/10.34172/ipp.2023.40574>
17. Ayyad M, Ayaad O, Alkhatatbeh H, Sawaqed F, Al-Rawashdeh S. Laparoscopic Partial Nephrectomy: Off-Clamp Versus on Clamp. *Asian Pacific journal of cancer prevention : APJCP*. 2022 05 01;23(5). <https://doi.org/10.31557/APJCP.2022.23.5.1719>
  18. Poppelen NM, Bruyn DP, Bicer T, Verdijk R, Naus N, Mensink H, Paridaens D, et al. Genetics of Ocular Melanoma: Insights into Genetics, Inheritance and Testing. *International Journal of Molecular Sciences*. 2020 Dec 30;22(1):336. <https://doi.org/10.3390/ijms22010336>
  19. You M, Xie Z, Zhang N, Zhang Y, Xiao D, Liu S, Zhuang W, et al. Signaling pathways in cancer metabolism: mechanisms and therapeutic targets. *Signal Transduction and Targeted Therapy*. 2023 05 10;8(1):196. <https://doi.org/10.1038/s41392-023-01442-3>
  20. Wang Q, Shao X, Zhang Y, Zhu M, Wang fxc, Mu J, Li J, et al. Role of tumor microenvironment in cancer progression and therapeutic strategy. *Cancer Medicine*. 2023 05;12(10):11149-11165. <https://doi.org/10.1002/cam4.5698>
  21. Nathan P, Hassel JC, Rutkowski P, Baurain J, Butler MO, Schlaak M, Sullivan, et al. Overall Survival Benefit with Tebentafusp in Metastatic Uveal Melanoma. *The New England Journal of Medicine*. 2021 09 23;385(13):1196-1206. <https://doi.org/10.1056/NEJMoa2103485>
  22. Parappilly MS, Chin Y, Whalen RM, Anderson AN, Robinson TS, Strgar L, Sutton TL, et al. Circulating Neoplastic-Immune Hybrid Cells Predict Metastatic Progression in Uveal Melanoma. *Cancers*. 2022 09 23;14(19):4617. <https://doi.org/10.3390/cancers14194617>
  23. Kurtenbach S, Sanchez mi, Kuznetsoff J, Rodriguez DA, Weich N, Dollar JJ, Cruz A, et al. PRAME induces genomic instability in uveal melanoma. *Oncogene*. 2024 02;43(8):555-565. <https://doi.org/10.1038/s41388-023-02887-0>
  24. Kaliki S, Shields CL. Uveal melanoma: relatively rare but deadly cancer. *Eye (London, England)*. 2017 02;31(2):241-257. <https://doi.org/10.1038/eye.2016.275>
  25. Solnik M, Paduszyńska N, Czarnecka am, Synoradzki kj, Yousef ya, Chorągiewicz T, Rejdak R, Toroet al. Imaging of Uveal Melanoma-Current Standard and Methods in Development. *Cancers*. 2022 06 27;14(13):3147. <https://doi.org/10.3390/cancers14133147>
  26. van Beek JGM, Koopmans AE, Verdijk RM, Naus NC, de Klein A, Kilic E. Diagnosis, histopathologic and genetic classification of uveal melanoma. In: Huynh Thien, D. (ed.) *Melanoma*. Rijeka: IntechOpen. Chapter 5. 2013;. <https://doi.org/10.5772/50853>
  27. Jiblawi A, Chanbour H, Tayba A, Khayat H, Jiblawi K. Magnetic Resonance Imaging Diagnosis of Choroidal Melanoma. *Cureus*. 2021 07;13(7):e16628. <https://doi.org/10.7759/cureus.16628>
  28. Bruyn DP, Beasley AB, Verdijk RM, Poppelen NM, Paridaens D, Keizer ROB, Naus NC, et al. Is Tissue Still the Issue? The Promise of Liquid Biopsy in Uveal Melanoma. *Biomedicines*. 2022 02 21;10(2):506. <https://doi.org/10.3390/biomedicines10020506>
  29. Park JJ, Diefenbach RJ, Byrne, Long GV, Scolyer RA, Gray ES, Carlino MS, Rizos H. Circulating Tumor DNA Reflects Uveal Melanoma Responses to Protein Kinase C Inhibition. *Cancers*. 2021 04 06;13(7):1740. <https://doi.org/10.3390/cancers13071740>
  30. Eleuteri A, Rola AC, Kalirai H, Hussain R, Sacco J, Damato BE, Heimann H, Coupland SE, Taktak AFG. Cost-utility analysis of a decade of liver screening for metastases using the Liverpool Uveal Melanoma Prognosticator Online (LUMPO). *Computers in Biology and Medicine*. 2021 03;130:104221. <https://doi.org/10.1016/j.combiomed.2021.104221>
  31. Uguen A. Digital Pathology Slides-based Measurement of Tumor Cells and Lymphocytes Within Cytology Samples Supports the Relevance of the Separation by Size of Nonhematological Tumor and Hematological Nontumor Cells in Liquid Biopsies. *Applied immunohistochemistry & molecular morphology: AIMM*. 2021 08 01;29(7):494-498. <https://doi.org/10.1097/PAI.0000000000000931>
  32. Bustamante P, Tsering T, Coblentz J, Mastromonaco C, Abdouh M, Fonseca C, Proença RP, et al. Circulating tumor DNA tracking through driver mutations as a liquid biopsy-based biomarker for uveal melanoma. *Journal of experimental & clinical cancer research: CR*. 2021 06 16;40(1):196. <https://doi.org/10.1186/s13046-021-01984-w>
  33. Armakolas A, Kotsari M, Koskinas J. Liquid Biopsies, Novel Approaches and Future Directions. *Cancers*. 2023 03 03;15(5):1579. <https://doi.org/10.3390/cancers15051579>
  34. Baum SH, Westekemper H, Bechrakis NE, Mohr C. Conjunctival and uveal melanoma: Survival and risk factors following orbital exenteration. *European Journal of Ophthalmology*. 2022 01;32(1):612-619. <https://doi.org/10.1177/1120672121995131>
  35. Branisteanu DC, Bogdanici CM, Branisteanu DE, Maranduca MA, Zemba M, Balta F, Branisteanu CI, Moraru AD. Uveal melanoma diagnosis and current treatment options (Review). *Experimental and Therapeutic Medicine*. 2021 Dec;22(6):1428. <https://doi.org/10.3892/etm.2021.10863>
  36. Shields CL, Shields JA. Enucleation for uveal melanoma. In: Albert, D.M., Miller, J.W., Azar, D.T., Young, L.H. (eds.) *Albert and Jakobiec's Principles and Practice of Ophthalmology*. Cham: Springer. 2022;;pp. 7717-7727. [https://doi.org/10.1007/978-3-319-90495-5\\_252-1](https://doi.org/10.1007/978-3-319-90495-5_252-1)
  37. Heng JS, Perzia BM, Sinard JH, Pointdujour-Lim R. Local recurrence of uveal melanoma and concomitant brain metastases associated with an activating telomerase promoter mutation seven years after secondary enucleation. *American Journal of Ophthalmology Case Reports*. 2022 09;27:101607. <https://doi.org/10.1016/j.ajoc.2022.101607>
  38. Negretti GS, Gurudas S, Gallo B, Damato B, Arora AK, Sivaprasad S, Sagoo MS. Survival analysis following enucleation for uveal melanoma. *Eye (London, England)*. 2022 08;36(8):1669-1674. <https://doi.org/10.1038/s41433-021-01710-y>
  39. Zemba M, Dumitrescu O, Gheorghe A, Radu M, Ionescu MA, Vatafu A, Dinu V. Ocular Complications of Radiotherapy in Uveal Melanoma. *Cancers*. 2023 01 04;15(2):333. <https://doi.org/10.3390/cancers15020333>
  40. Yamada K, Takeuchi M, Fukumoto T, Suzuki M, Kato A, Mizuki Y, Yamada N, et al. Immune checkpoint inhibitors for metastatic uveal melanoma: a meta-analysis. *Scientific Reports*. 2024 04 03;14(1):7887. <https://doi.org/10.1038/s41598-024-55675-5>
  41. Brănișteanu DE, Porumb-Andrese E, Porumb V, Stărică A, Moraru AD, Nicolescu AC, Zemba M, et al. New Treatment Horizons in Uveal and Cutaneous Melanoma. *Life (Basel, Switzerland)*. 2023 07 31;13(8):1666. <https://doi.org/10.3390/life13081666>
  42. Smith KER, Peng K, Pulido JS, Weisbrod AJ, Strand CA, Allred JB, Newsom AN, et al. A phase I oncolytic virus trial with vesicular stomatitis virus expressing human interferon beta and tyrosinase related protein 1



- administered intratumorally and intravenously in uveal melanoma: safety, efficacy, and T cell responses. *Frontiers in Immunology*. 2023;14:1279387. <https://doi.org/10.3389/fimmu.2023.1279387>
43. Gezgin G, Visser M, Ruano D, Santeagoets SJ, Miranda NFCC, Velden PA, Luyten GPM, et al. Tumor-Infiltrating T Cells Can Be Expanded Successfully from Primary Uveal Melanoma after Separation from Their Tumor Environment. *Ophthalmology Science*. 2022 06;2(2):100132. <https://doi.org/10.1016/j.xops.2022.100132>
  44. Barbi M, Carvajal RD, Devoe CE. Updates in the Management of Uveal Melanoma. *Cancer Journal (Sudbury, Mass.)*. 2024 04 01;30(2):92-101. <https://doi.org/10.1097/PPO.0000000000000708>
  45. BeigiYZ, Lanjanian H, Fayazi R, Salimi M, Hoseyni BHM, Noroozizadeh MH, Masoudi-Nejad A. Heterogeneity and molecular landscape of melanoma: implications for targeted therapy. *Molecular Biomedicine*. 2024 05 10;5(1):17. <https://doi.org/10.1186/s43556-024-00182-2>
  46. Wu YY, Sun TK, Chen MS, Munir M, Liu H. Oncolytic viruses-modulated immunogenic cell death, apoptosis and autophagy linking to virotherapy and cancer immune response. *Frontiers in Cellular and Infection Microbiology*. 2023;13:1142172. <https://doi.org/10.3389/fcimb.2023.1142172>
  47. Demirci H, Narvekar A, Murray C, Rich C. A phase II trial of AU-011, an investigational, virus-like drug conjugate (VDC) for the treatment of primary indeterminate lesions and small choroidal melanoma (IL/CM) using suprachoroidal administration. *Annals of Oncology*. 2022;33(Suppl):S934. Available from: [https://www.annalsofncology.org/article/S0923-7534\(22\)02819-8/fulltext](https://www.annalsofncology.org/article/S0923-7534(22)02819-8/fulltext)
  48. Wu F, Lane AM, Trofimov A, Shih HA, Gragoudas ES, Kim IK. Outcomes after Proton Beam Irradiation in Patients with Choroidal Melanoma Eligible for Investigational AU-011 Treatment. *Ocular Oncology and Pathology*. 2023 Dec;9(5-6):152-157. <https://doi.org/10.1159/000534184>
  49. Alnefaie A, Albogami S, Asiri Y, Ahmad T, Alotaibi SS, Al-Sanea MM, Althobaiti H. Chimeric Antigen Receptor T-Cells: An Overview of Concepts, Applications, Limitations, and Proposed Solutions. *Frontiers in Bioengineering and Biotechnology*. 2022;10:797440. <https://doi.org/10.3389/fbioe.2022.797440>
  50. Jakubecova J, Smolkova B, Furdova A, Demkova L, Altanerova U, Nicodemou A, Zeleznikova T, et al. Suicide-Gene-Modified Extracellular Vesicles of Human Primary Uveal Melanoma in Future Therapies. *International Journal of Molecular Sciences*. 2023 08 19;24(16):12957. <https://doi.org/10.3390/ijms241612957>
  51. Milán-Rois P, Quan A, Slack FJ, Somoza Á. The Role of LncRNAs in Uveal Melanoma. *Cancers*. 2021 08 11;13(16):4041. <https://doi.org/10.3390/cancers13164041>
  52. Barbagallo C, Di Maria A, Alecci A, Barbagallo D, Alaimo S, Colarossi L, Ferro A, et al. VECTOR: An Integrated Correlation Network Database for the Identification of CeRNA Axes in Uveal Melanoma. *Genes*. 2021 06 29;12(7):1004. <https://doi.org/10.3390/genes12071004>
  53. Yang C, Wang R, Hardy P. Potential of miRNA-Based Nanotherapeutics for Uveal Melanoma. *Cancers*. 2021 Oct 16;13(20):5192. <https://doi.org/10.3390/cancers13205192>
  54. Majed M, Ayaad O, AlHasni NS, Ibrahim R, AlHarthy SH, Hassan KK, Al-Zadiali R, Al-Awaisi H, Al-Baimani K. Reducing the Risk of Fall among Oncology Patients using Failure Modes and Effects Analysis. *Asian Pacific journal of cancer prevention: APJCP*. 2024 02 01;25(2):689-697. <https://doi.org/10.31557/APJCP.2024.25.2.689>
  55. AlHarthy SH, Ayaad O, Al Mashari AAA, AlBalushi MA, Ibrahim R, Bait Nasib MH, Al Zadiali R, Awaisi H, Al Baimani K. Improving Care Continuity in Oncology Settings: A Lean Management Approach to Minimize Discharges Without Follow-Up Appointments. *Asian Pacific journal of cancer prevention: APJCP*. 2024 04 01;25(4):1293-1300. <https://doi.org/10.31557/APJCP.2024.25.4.1293>
  56. Al-Ruzzieh MA, Al-Helih YM, Ayaad O, Haroun A, Alnaimat S. Comprehensive evaluation of patient-centered care at cancer center: A qualitative descriptive study. *Nursing Forum*. 2025 02 23;:5070345. <https://doi.org/10.1155/nuf/5070345>
  57. Al-Ruzzieh MA, Al-Helih YM, Haroun A, Ayaad O. Higher and Middle Management Perspectives on Patient-Centered Care in an Oncology Setting: A Qualitative Study. *Nursing Reports (Pavia, Italy)*. 2024 Nov 05;14(4):3378-3390. <https://doi.org/10.3390/nursrep14040244>
  58. Ayaad O, Ibrahim R, AlHasni NS, Salman BM, Sawaya ZC, Zadiali RA, Faliti BA, et al. Assessing Health Literacy, Learning Needs, and Patient Satisfaction in Cancer Care: A Holistic Study in the Omani Context. *Asian Pacific Journal of Cancer Biology*. 2024 Nov 23;9(4):553-560. <https://doi.org/10.31557/apjcb.2024.9.4.553-560>
  59. Ayaad O, Ibrahim R, AlBaimani K, AlGhaithi MM, Sawaya ZG, AlHasni NS, AlAwaisi HS, et al. Predicting and Classifying the Perceptions of Learning Needs Importance in Cancer Patients; a Machine Learning Approach. *Health Education and Health Promotion*. 2024 Oct 10;12(4):649-660. <https://doi.org/10.58209/hehp.12.4.649>
  60. Ayyad M, Ayaad O. Measuring quality of life among patients with urinary stone disease; A qualitative study. *Journal of Renal Injury Prevention*. 2022 09 18;12(4):e32106-e32106. <https://doi.org/10.34172/jrip.2023.32106>
  61. Salman BM, Ayaad O, Ibrahim R, AlHartrushi MS, Majed M, Al Zadiali R, AlTobi ZA, et al. Enhancing Medication Safety: Reducing Administration Errors in Oncology Setting. *Asian Pacific journal of cancer prevention: APJCP*. 2025 01 01;26(1):269-277. <https://doi.org/10.31557/APJCP.2025.26.1.269>
  62. AlSheidi SA, Ayaad O, Ibrahim R, AlDahli SN, Majed M, AlWaheibi HM, Zadiali ROAA, et al. Optimizing Laboratory Processes: A Path to Reduced Sample Rejection in Oncology. *Iranian Journal of Public Health*. 2025 01;54(1):155-165. <https://doi.org/10.18502/ijph.v54i1.17587>
  63. Al Qassabi B, Al Sukaiti R, Alajmi S, Sheikh Omar A, Ibrahim R, Al Faliti B, Zribi A, et al. Improving the Timely Reporting of Critical Radiological Results in Oncology to Enhance Patient Safety (A Quality Improvement Initiative at SQCCRC). *Asian Pacific journal of cancer prevention: APJCP*. 2025 03 01;26(3):1089-1097. <https://doi.org/10.31557/APJCP.2025.26.3.1089>
  64. Haddabi IHA L, Ibrahim R, AlSheidi SA, Busaidi A, Ghufuran N, AlDahli SN, Awor OA, et al. Minimizing the Risk of Sample Mix-ups in the Molecular Pathology Section in Oncology Center Using Risk Assessment Matrix (RAM). *Asian Pacific Journal of Cancer Biology*. 2025 01 12;10(1):37-45. <https://doi.org/10.31557/apjcb.2025.10.1.37-45>
  65. AlHarthy S, Al-Moundhri M, Al-Mahmoodi W, Ibrahim R, Ayaad O, Al Baimani K. Referral Process Enhancement: Innovative Approaches and Best Practices. *Asian Pacific journal of cancer prevention: APJCP*. 2024 05 01;25(5):1691-1698. <https://doi.org/10.31557/APJCP.2024.25.5.1691>
  66. Al Qassabi B, AlSukaiti R, Alajmi S, Omar AS, Ibrahim R, Banibakr AA, Al-Baimani K, et al. Improving Turnaround Times and Operational Efficiency in Radiology Services: Quality Improvement Study in Oman. *Asian Pacific journal*

- of cancer prevention: APJCP. 2025 05 01;26(5):1709-1718. <https://doi.org/10.31557/APJCP.2025.26.5.1709>
67. Ayaad O, Al-Dewiri R, Kasht L, Qaddumi B, Ayyad M. Adopting Lean Management in Quality of Services, Cost Containment, and Time Management. Asian Pacific journal of cancer prevention: APJCP. 2022 08 01;23(8):2835-2842. <https://doi.org/10.31557/APJCP.2022.23.8.2835>
  68. Haroun A, Ayaad O, Al-Ruzzieh MA, Ayyad M. Role of Total Quality Management in patient experience. British Journal of Healthcare Management. 2022;28(10):1-8. <https://doi.org/10.12968/bjhc.2021.0082>
  69. Al-Ruzzieh MA, Al-Helih YM, Ayaad O, Hess RG. The Influence of Emotional Intelligence on Shared Governance Councils Effectiveness Among Nurses Participating in Shared Governance Councils in an Oncology Setting. The Journal of Nursing Administration. 2025 03 01;55(3):172-176. <https://doi.org/10.1097/NNA.0000000000001552>
  70. Al-Ruzzieh MA, Eddin R, Ayaad O, Kharabsheh M, Al-Abdallah D. Examining Nurse and Patient Factors Before and After Implementing an Oncology Acuity Tool: A Mixed Methods Study. Journal of Nursing Measurement. 2024 03 14;32(1):38-46. <https://doi.org/10.1891/JNM-2022-0001>
  71. Al-Ruzzieh MA, Ayaad O. Nurses' emotional intelligence and professional practice models. British Journal of Nursing. 30(19):1110-6. <https://doi.org/10.12968/bjon.2021.30.19.1110>
  72. Ayaad O, Alloubani A, Thiab F, Yousef D, Banat B. Shared governance to improve nursing environments. British Journal of Healthcare Management. 2018;24(12):594-602. <https://doi.org/10.12968/bjhc.2018.24.12.594>
  73. Abuseif S, Ayaad O, Abu-Al-Haijaa E. Factors affecting nurse autonomy. International Journal of Academic Research in Business and Social Sciences. 2018;8(12):1785-96. <https://doi.org/10.6007/IJARBS/v8-i12/5323>
  74. Al-Ruzzieh MA, Ayaad O. Nursing Professional Practice Model: Development, Implementation, and Evaluation at an International Specialized Cancer Center. The Journal of Nursing Administration. 2020 Nov;50(11):562-564. <https://doi.org/10.1097/NNA.0000000000000937>
  75. Qaddumi B, Ayaad O, Al-Ma'aitah MA, Akhu-Zaheya L, Alloubani A. Team effectiveness and collaborative tools. Journal of Interprofessional Education & Practice. 2021;24:100449. <https://doi.org/10.1016/j.xjep.2021.100449>
  76. Al-Ruzzieh MA, Al Rifai A, Ayaad O. Organizational citizenship in healthcare: A scoping review. British Journal of Healthcare Management. 2022;28(6):1-7. <https://doi.org/10.12968/bjhc.2021.0039>
  77. AlHasni NS, Ayaad O, Al-Awaisi HS, Ibrahim RA, Al Faliti BHS, AlMadhoun ET, Al-Baimani K. Correlation between Innovation Practices and Occupational Fatigue in Healthcare Professionals. Health Education and Health Promotion. 2025 01 10;13(1):13-19. <https://doi.org/10.58209/hehp.13.1.13>
  78. Ayaad O, Al-Ruzzieh MA, Qaddumi B, Al Hroub A, Ayyad M, Abuseif S, Çelik Y. Outsourcing services in healthcare. British Journal of Healthcare Management. 28(3):96-103. <https://doi.org/10.12968/bjhc.2020.0171>



This work is licensed under a Creative Commons Attribution-Non Commercial 4.0 International License.